**Disk Partition Techniques Assessment and Analysis Applied to Genomic Assemblers Based on Bruijn Graphs**

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**Abstract:** In this paper an assessment of several de-novo genomic assembler tools based on de Bruijn graph is made, with the purpose to measure the impact of the use of disk partitioning techniques regarding the computational requirements and generate a framework for bioinformatics researchers to let them identify advantages, disadvantages, bottlenecks and challenges of the assemblers using those techniques.

Assessed assemblers using disk partitioning techniques were: Minia and EPGA, the assessed assemblers that do not use disk partitioning were: ABySS and SOAPDenovo2. The parameters measured were the following: occupied space in RAM, processing time, parallelization and disk read and write access. A dataset was used with 36,504,800 short reads corresponding to 14th human chromosome. The assessment was made for two kmer sizes: 31 and 55. The results obtained were the following: The tools based on disk partitioning techniques showed the less RAM use. The tools with more I/O transfer intensity were the ones using disk partitioning techniques. The techniques that achieved more parallelization were the ones using disk partitioning.

**Key words:** Assemblers, assembly, bioinformatics, kmer count, minimizers.

**1. Introduction**

The rapid fall of sequencing costs since the arrival of the next generation sequencing (NGS) allowed the possibility to sequence complete genomes of different organisms with the aim to ease the study of their genes. One of the main problems and challenges is in the de-novo assembly stage [1], [2], in this stage short reads are used (between 100bp and 500bp) from machines with Illumina or Roche 454 technology and are assembled in contigs and metacontigs in such a way that as a result a complete assembled genome is obtained without need to count with a reference genome. For the assembly of small genomes, such as the bacterial genome assembly, these tools have good performance and most of the time require a little computation time. However as the genome size increases (e.g. the chromosome of a mammal) the computing requirements increases to perform this process [3].

Computational intensity required to perform an assembly without a reference genome (de-novo) has been addressed in the last decade using techniques based on De Bruijn graphs [4]. Actually, the most used genomic assemblers are based on this approach, such as Velvet [5], ALLPATHS [6], ABySS [7], SOAPdenovo2 [8], MINIA [9].
Despite the remarkable results in the reduction of computing time for the assemblers based on De Bruijn, one of the aspects where high demand is still present is the memory use, mainly due to the generation, storage and kmer analysis and the graph representation. To mitigate these high memory requirement presented, in the last 5 years there have been assembler implementations that use disk partitioning techniques in some of its stages. Below is described the main techniques of disk partitioning used in the genomic assembly.

1.1. Kmers Distribution through a Hash Function

DSK [10] uses this method to make kmers partitioning in disk. To make the kmers disk partitioning is necessary to calculate the number of kmers, the number of iterations (ni) to cycle through the kmers and the number of partitions (np) to be created in disk. To make this calculation it must have input parameters such as the maximum memory use M (bits) and the kmer size k.

The hash function used in this algorithm is a function (h) that maps a kmer to a numeric value between [0 and H] where H is a big integer (2^64).

For each iteration, kmers (m) are taken one by one from the sequence. For each kmer the hash is calculated and the module operation is executed against the number of iterations (h(m) mod ni), if the result of this operation is equal to the number of the actual iteration (i) the kmer is written to a file Dj where j is h(m)/ni mod np.

This method reduce significantly reduces the use of RAM in the kmers counting stage because it is not necessary to load all the kmers in memory. However, it may be the case that the kmers are not evenly distributed across all partitions.

1.2. Minimizers

The concept of minimizers was introduced for the first time in 2004 [11] with the purpose of facilitate the comparison of sequences because the method seed-and-extend is usually used for this task, this method requires a seed catalog that could take large amount of memory in the case of mammalian genomes or large proteins. To reduce the necessary space is required to save less kmers. The proposed method in [11] consist in choosing a representative kmer from a group of adjacent kmers such that different strings of text Ti and Tj have the same representative kmer if they share a large subsequence. A minimizer is a sub-chain (fixed size less than k) of a kmer with less lexicographical weight. However in a recent paper [12] the lexicographical weight is replaced by the frequency of the possible minimizers allowing to distribute more evenly the kmers on partitions.

In the case of assembly, inBCALM [12] the l-minimizer of a string u is the smallest l-mer that is a sub-string of u (assuming there is a total ordering of the strings, e.g. lexicographical). They define Lmin(u) (respectively, Rmin(u)) to be the l-minimizer of the (k − 1)-prefix (respectively suffix) of u. They refer to these as the left and right minimizers of u, respectively. They therefore perform in-memory l-mer counting to obtain a sorted frequency table of all l-mers. This step requires an array of 64|Σ| l bits to store the count of each l-mer in 64 bits, which is negligible memory overhead for small values of l (8 MB for l = 10). Each l-mer is then mapped to its rank in the frequency array, to create a total ordering of minimizers.

In previous works such as Assemblathon 1[13] Assemblathon 2 [14], and Genome Assembly Gold-Standard Evaluations (GAGE) [15] performance tests were conducted to the assemblers regarding to their results, that is, the number of generated contigs and their statistics. However in these works few mentions or none about the variables such as read/write operations, the RAM use and the parallelization of these assemblers. Moreover, some of the performance assessments available focus in the general performance analysis without focusing on each of its stages.

In this paper is proposed to assess several genomic assemblers based on De Bruijn graphs in order to
compare the computational demand of those that use disk partitioning techniques with respect to those that do not use them. For each assessed tool the data related to writing and reading, main memory use (RAM), parallelization (number and percentage of used processors) and processing time is measured with the purpose to verify which tools and which stages has major computational requirements.

2. Materials and Methods

2.1. Datasets

The dataset used to this assessment corresponds to 64587949 short reads (101bp), paired-end of the 14th Homo Sapiens chromosome.

2.2. Computational Equipment

The computer where this assessment was in place has the following described characteristics: Operating System: Debian Wheezy (AMD64). Processor: Intel(R) Xeon(R) cpu E7450 @ 2.40GHz, 24 cores. RAM: 64GB. Hard Disk: 160GB HDD.

2.3. Assessed Assemblers

The tools to evaluate are assemblers based on de bruijn graphs that use only short paired-end reads for contigs determination. Assemblers were chosen in such a way that there was diversity of kmercounting algorithms and the graph representation structures in memory, additionally as a selection criterion was taken the major use during the last decade and its lastest version was published in the year were that assessment was performed (2015). The tools to evaluate were grouped according to the use or not of disk partitioning techniques. Note that assemblers such as ALL-PATHS-LG were not included in this assessment because it requires long fragments as additional to the short fragments to be executed.

2.3.1. Assessed assemblers that does not use disk partitioning techniques

ABySS: Is a de novo parallel assembler for paired sequences, specifically designed for short reads. Roughly the algorithm used by this tool has the following stages: First, without using the paired-end information, contigs are extended until either they cannot be unambiguously extended or come to a blunt end due to a lack of coverage. In the second step the paired-end information is used to resolve ambiguities and merge contigs [7]. ABySS is implemented in C++ and use the Openmpi library (in some of its stages) with the purpose to allow the communication between several computer nodes and ease its execution on a cluster. Additionally ABySS use the google-sparsehash library to decrease the RAM requirement in the kmers counting.

SOAPdenovo2: Is a novel short-read assembly method that can build a de novo draft assembly for the human-sized genomes. This algorithm has the following steps: De Bruijn Graph (DBG) construction, contig assembly, paired-end (PE) reads mapping, scaffold construction, and gap closure. SOAPdenovo2 is implemented in C++ and for some of its processing stages use several threads.

2.3.2. Assessed assemblers that use disk partitioning techniques

Minia: Is a short-read assembler based on a de Bruijn graph, capable of assembling a human genome on a desktop computer in a day. The output of Minia is a set of contigs. The main stages for the Minia algorithm are: kmer counting, graph construction and the cycle or simplification of it. For the kmer counting stage, Minia uses DSK that use the distribution on disk of kmers method through a hash function with the purpose to reduce the RAM memory use. Minia is implemented on C++. Note that as opposed to Abyss, Minia does not use the available paired-end information, this means that it does not create metacontigs.

EPGA2: Is an genomic assembler oriented to efficient memory use. To accomplish this purpose it uses BLESS [16] for error correction in the reads, the kmers distribution in disk method through the hash...
function with the use of DSK [10] for the kmer counting stage and the minimizers method through BCALM [12] for node simplification in the De Bruijn graph. Once the nodes are simplified and contigs generated EPGA2 joins the contigs in parallel [17].

3. Results

The Fig. 1 and the Fig. 2 show the RAM use a peak for each assembler and its stages, for parameters $k=31$ and $k=55$ respectively.

![Fig. 1. RAM use peaks for each tool ($k=31$).](image1)

![Fig. 2. RAM use peaks for each tool ($k=55$).](image2)

The Fig. 3 and Fig. 4 show the CPU usage number peak and average for each assembler and its stages for parameters $k=31$ and $k=55$ respectively.
Fig. 3. CPUs use peaks for each tool ($k=31$).

Fig. 4. CPUs use peaks and averages for each tool ($k=55$).

Fig. 5. Peaks and average I/O transfer for each tool ($k=55$).
The Fig. 5 and Fig. 6 show the peak and average I/O transfer for each assembler's stages and its stages for the parameter $k=31$ and $k=55$ respectively.

Fig. 6. Peaks and average I/O transfer for each tool ($k=55$).

Fig. 7. Execution time of each tool ($k=31$).

Fig. 8. Execution time of each tool ($k=55$).
Fig. 7 and Fig. 8 show the execution time used by each stage for each assembler for the parameters $k=31$ and $k=55$ respectively.


Regarding the RAM use it can be concluded that for the counting kmers stage the tools with the less RAM use were the ones using disk partitioning techniques through a hash function, for example, Minia and EPGA2 using DSK. The tool with the less RAM use in the contigs generation stage was EPGA, since it makes use of disk partitioning by minimizers technique (selected by frequency) in the stage of contigs generation (BCALM).

Regarding the I/O transfer it was noted that the tools that use disk partitioning techniques had the highest average of transfer in the counting kmer stage. Additionally, for the contigs generation stage the tool with the highest average I/O transfer was EPGA, because of the use of BCALM to simply nodes in the graph.

As for parallelization it can be noted that the tools that use disk partitioning have greater parallelization in the counting kmers stage.

Regarding the execution time, the tools that use disk partitioning took the less execution time in the counting kmers stage, however in the metacontigs generation stage EPGA required a greater amount of time.

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References


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