

Potential Activity of the *Achillea wilhelmsii* Leaves on Bacteria

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Abstract—*Achillea wilhelmsii* is belong to *Asteraceae* family. This plant is from medicinal plants in Iran customary medicine. The aim of this study was to appraise of effects of the leaves essential oil and methanol extract of the *Achillea wilhelmsii* on the growth of the bacteria. In this study after collecting and provision plant, leaves essential oil were obtained by hydrodistillation and leaves methanol extract were obtained using a Soxhlet apparatus. The effects of methanol extract antimicrobial were assessed using Agar Well Diffusion method and also using Dilution Test method. The effects of essential oil antimicrobial were determined by using Agar Dilution Assay. Dates were analyzed using Chi-square and AVONA test in the $p < 0/001$. Leaves methanol extract had more effect against *Bacillus cereus* and *Staphylococcus aureus* bacteria and showed weak effect against *Escherchia coli* and was not observed any growth inhibition effect against *Pseudomonas aeruginosa*, also leaves essential oil had inhibition effect against *Bacillus cereus*, *Staphylococcus aureus* and *Escherchia coli* and did not have any inhibition effect against *Pseudomonas aeruginosa*. Leaves methanol extract and essential oil of *Achillea wilhelmsii* have antibacterial effects, therefore we will be able perform researches with extraction of this plant effective compound for the treatment of infectious diseases.

Keywords—*Achillea wilhelmsii*, antibacterial effect, essential oil, methanol extract.

I. INTRODUCTION

Use of traditional medicine in Asia a long history of human interactions with the environment. Plants used for traditional medicine contain substances that can be used to treat chronic as well as infectious diseases. A knowledge of how to use the plants against different illnesses may be expected to have accumulated in areas where the use of plants is still of great importance. The medicinal worth of plants lies in some chemical substances that produce a definite physiological action on the human body. The most important of these bioactive compounds of plants are tannins, alkaloids, flavanoids and phenolic compounds [1]. In developing countries, farmers, people of small isolate villages and native communities use folk medicine for the treatment of common infections. The *Achillea*, which belongs to the family Compositae (*Asteraceae*) and comprises more than 120

Manuscript received August 16, 2011; revised September 1, 2011.

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species. These plants are medicinal perennial herbs that are native to Europe and Western Asia, although they are also found in Australia, New Zealand and North America [2]. Several effects, such as anti-inflammatory, antihypertensive, and anti-hyperlipidemia and antitumor have been reported for *Achillea*. It is widely used in traditional medicine for gastrointestinal disorders and there are some reports of its effects, such as antispasmodic, choleric, antiulcer, antibacterial (*Helicobacter pylori*), and hepatoprotective, on the gastrointestinal tract [3]. *Achillea wilhelmsii* C. Koch (*Asteraceae*) is widely found in different parts of Iran. This plant is full of flavonoids and sesquiterpene lactones, which have been shown to be effective in lowering blood lipids and hypertension [4], and widely used in Iranian traditional medicine for gastrointestinal disorders. It has chemical components, including flavonoids, alkaloids (achilleine), cineol, borneol, α - and β -pinen, camphor, caryophyllene, thujene, rutin, sesquiterpenoids, and monoterpenoids [3].

Thus, the main aim of the present project was to carry out a biological investigation on *Achillea wilhelmsii* C. Koch from the Iran. In this study, *Achillea wilhelmsii* were collected for evaluation of their antimicrobial activities and were tested against several pathogenic bacteria.

II. MATERIAL AND METHODS

Achillea wilhelmsii C. Koch was collected from around area of Isfahan city (Iran) in the May of 2009. The voucher specimen was deposited at the herbarium of Research Institute of Isfahan Forests and Rangelands. The plant materials were dried under shade and ground into fine powder using electric blender, then, 60 g of leaves powder were extracted with 300 ml methanol (CH_3OH) by Soxhlet extraction for 8 hours. The residue was dried over night and then evaporated by using a rotary evaporator and freeze dryer. The dried extracts were stored at -20°C until used. The extracts were dissolved in 5% aqueous dimethylsulfoxide (DMSO) with and sterilized by filtration through a $0.45\ \mu\text{m}$ membrane filter. The essential oils were obtained by hydrodistillation using a Clevenger-type apparatus for 3 h, from leaves of *A. wilhelmsii*. The oil yields were $80\ \mu\text{l}$ (300g leaf powder with 1000 ml water). The essential oils were dissolved in 5% aqueous dimethylsulfoxide (DMSO) and were dried over anhydrous sodium sulfate overnight and kept in sterile sample tubes in refrigerator. Microorganisms were obtained from the Institute of Scientific and Industrial Researchs, Iran. Two strains of gram-negative bacteria *Escherichia coli* (ATCC 25922), *Pseudomonas aeruginosa* (ATCC 27853), and two strains of gram-positive bacteria *Bacillus cereus* (ATCC 1274) and *Staphylococcus aureus*

(ATCC 25923)] were used. The cultures of bacteria were maintained in their appropriate agar slants at 4°C throughout the study and used as stock cultures.

For the study of antibacterial activity, total count of isolated strains were standardized to equivalent a 0.5 MacFarland Nephelometer standard (1×10^8 cfu/ml) by Mueller-Hinton broth and then diluted as 1:10. Antibacterial activity was studied by the agar well diffusion method [5]. Mueller Hinton agar was used as the bacteriological medium. The extracts were diluted in 5% dimethylsulphoxide (DMSO) at the concentrations of 20 mg/ml, 30 mg/ml, 50 mg/ml and 400 mg/ml. The Mueller Hinton agar was melted and cooled to 48 – 50°C and a standardized inoculum (1.5×10^8 CFU/mL, 0.5 McFarland) was then added aseptically to the molten agar and poured into sterile Petri dishes to give a solid plate. Wells were prepared in the seeded agar plates. The test compound (100 µl) was introduced in the well (6 mm). The plates were incubated overnight at 37°C. The antimicrobial spectrum of the extract was determined for the bacterial species in terms of zone sizes around each well. The diameters of zone of inhibition produced by the agent were compared with those produced by the commercial control antibiotics, chloramphenicol (30 µl). The experiment was performed three times to minimize the error and the mean values are presented.

Minimal inhibitory concentration (MIC) and minimal bactericidal concentration (MBC) were determined for the extracts that showed total growth inhibition using the protocol described above. Extract concentrations of 6/25 to 200 mg/ml were evaluated. The concentration at which there was no visually detectable bacterial growth was taken as the MIC and the concentration at which there was no bacterial growth after inoculation in Mueller-Hinton agar was taken as the MBC [5]. Means of triplicate measurements and standard errors were determined for each sample.

III. RESULTS

Results were analyzed and bacteria growth inhibition were confirmed after inoculation in Mueller-Hinton agar. The extracts showed some degree of inhibition of bacterial growth at different concentrations. The methanol extract of the leaf exhibited antibacterial activity at varied levels against *Staphylococcus aureus*, *Bacillus cereus* and *Escherichia coli*. The methanol extract of *Achillea wilhelmsii* was least active against *Pseudomonas aeruginosa* in comparison to all the microorganisms tested. The extract was more active against Gram-negative bacteria as compared to Gram-positive. The inhibitory effect of the extract was compared with standard antibiotic, chloramphenicol.

The antibacterial activity of the methanol extract of the leaves *Achillea wilhelmsii* were determined against four bacterial strains which is reported in Table 1. The MIC and MBC of these extracts are currently being obtained and are reported in Table 2.

The essential oil of the leaf exhibited antibacterial activity at against *Staphylococcus aureus*, *Bacillus cereus* and *Escherichia coli*, but, has not antibacterial activity at against *Pseudomonas aeruginosa*.

TABLE I: ZONE OF INHIBITION (LEAVE METHANOLIC EXTRACT) FOR BACTERIA

Concentration	400 (mg/ml)	50 (mg/ml)	30 (mg/ml)	20 (mg/ml)	Control (-)	Control (+)
Bacteria						
<i>Staphylococcus aureus</i>	18± 0.816	18± 0.816	9± 0.816	8± 0.816	6	20
<i>Bacillus cereus</i>	21.75± 0.5	12	12	9± 0.816	6	18
<i>Escherichia coli</i>	9± 0.816	6	6	6	6	25
<i>Pseudomonas aeruginosa</i>	6	6	6	6	6	23

TABLE II: DETERMINATION OF MIC AND MBC FOR BACTERIA

Bacteria	MBC (mg/ml)	MIC (mg/ml)
<i>Staphylococcus aureus</i>	25	12.5
<i>Bacillus cereus</i>	12.5	6.25
<i>Escherichia coli</i>	200	100
<i>Pseudomonas aeruginosa</i>	-	-

MIC: Minimum Inhibitory Concentration

MBC: Minimum Bactericidal Concentration

IV. DISCUSSION

Plants have an extremely ability to synthesize aromatic products, most of which are phenols [6]. Most are secondary metabolites and in many cases, these products serve as plant defense mechanisms against predation by microorganisms, insects, and so forth. Some, such as terpenoids, give plants their odours, others (quinones and tannins) are responsible for plant pigment. Many compounds are responsible for plant flavor and some of the same herbs and spices used by humans to season food yield useful medicinal compounds [6].

Plant essential oils and extracts have been used in food preservation, pharmaceuticals, alternative medicine and natural therapies [7]. These are potential sources of novel antimicrobial compounds especially against bacterial pathogens. *Achillea wilhelmsii* is known for many years in the folk medicine. *In vitro* studies in this work showed that *Achillea wilhelmsii* leaves inhibited bacterial growth but their effectiveness varied. Totally, the Gram-positive bacteria are more susceptible than Gram-negative bacteria due to the differences in their cell wall structure. Gram-negative organisms are considered to be more resistant due to their outer membrane acting as a barrier to many environmental substances, including antibiotics [8]-[10]. However, the results from this study reveals that the essential oils of *Achillea wilhelmsii* leaves rich in monoterpenes, including: camphor, 1, 8 - cineole, borneol and myrtenol [11]. This compounds have been reported to display strong antibacterial effects [7], [12]-[13]. Thus, this plant is full of flavonoids and sesquiterpene lactones [4] and leaves methanolic extract have also been shown to possess good anti-bacterial activity. The studies carried out showed that the flavonoids and phenolic compounds have also exhibited notable antimicrobial activity as well [14]-[17]. In general, the essential oil showed better activity than the methanol extract. This observation agrees

with previous reports by Vukovic et al. (2007) and Sokmen et al. (2004) that methanol extracts showed less activity than essential oil [10],[12].

V. CONCLUSION

The results of this study indicate the antimicrobial activity of methanolic extract and essential oil of *Achillea wilhelmsii* leaves. Our results further support the idea that medicinal plants can be promising sources of potential antimicrobial agents. These results form a good basis for selection of the plant for further phytochemical and pharmacological investigation. However, more studies are needed to investigate the antimicrobial activity of flavonoids and terpenes compound kind in this plant and their potential for use in clinical situations.

Studies aimed at the isolation and structure elucidation of antibacterial active constituents from this plant is in progress.

ACKNOWLEDGMENT

This work was supported by Young Researchers Club that depended on Islamic Azad University, Falavarjan Branch, Iran. The authors also thank Dr. Monajjemi and Ms. Emami from the Department of Biology, Islamic Azad University, Falavarjan Branch for their aid.

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