# Bovine STAT5A Gene Polymorphism Analysis and Its Association with Milk Composition Traits in Jersey Cows

M. Selvaggi, V. Tufarelli, F. Pinto, G. Centoducati, A. Dambrosio, M. P. Santacroce, and C. Dario

Abstract—In mammals, the STAT proteins (signal transducers and activators of transcription) are a family of cytoplasmic transcription factors mediating the actions of many peptide hormones and cytokines within target cells. In particular, STAT5A is a crucial mediator in the lactogenic hormone response being a candidate marker for milk traits in farm animals. In the present paper, the  $T \rightarrow C$  nucleotide polymorphism at position 12743 in exon 16 of the bovine STAT5A gene was analyzed with PCR-RFLP in a sample of Jersey cows. The purposes of this investigation were to determine the frequencies of the variant alleles and the genotypes of this SNP in Jersey cows and to verify its association with some milk production traits. All the three possible genotypes were identified in the studied population. The observed frequencies of C and T alleles were 0.147 and 0.853 repectively. The TT genotype was the most frequent followed by TC and CC ones. No significant differences between the TT and TC genotypes were found considering MY, FY PC and PY. On the other side, the difference concerning the fat content of milk produced by cows belonging to TC and TT groups was found significant at the statistical analysis: in particular, milk from TT animals had a higher fat content in comparison with that of TC ones (4.55 vs. 4.14%, respectively; P < 0.05). However it may be necessary to carry out further investigations about this SNP to better clarify its role on milk production traits in cattle.

*Index Terms*—Jersey breed, milk production, SNP, STAT5A gene.

### I. INTRODUCTION

The recent advances of molecular genetics in the identification of loci affecting production traits of domestic animals have opened new and interesting opportunity for their genetic improvement. With the use of molecular technologies, it may be possible to select a breeding animal for a wide range of traits and to enhance reliability in predicting the mature phenotype of the individual. Milk production traits, which are under control of several genes, are very important in dairy cattle due to their economic relevance: to improve milk yield and composition is of great significance for breeders.

The STATs proteins (signal transducers and activators of

transcription) comprise a 7-member family of latent cytoplasmic transcription factors that mediate actions of many peptide hormones and cytokines within target cells [1], [2]. They act as signal transducers in the cytoplasm and transcription activators in the nucleus. The DNA-binding capacity of STATs is induced by phosphorylation of a tyrosine residue at the C-terminus of the protein, which leads to dimerization and nuclear localization, and this has been shown to occur in response to a wide range of hormones and cytokines. STAT5 was initially discovered as a PRL-induced transcription factor and named mammary gland factor (MGF) [3]. STAT5 is also known as a main mediator of growth hormone (GH) action on target genes [4]; it is a key intracellular mediator of prolactin signalling and can activate transcription of milk protein genes in response to prolactin [3]. Initially, a single STAT5 gene was identified in sheep but subsequently two forms of STAT5 (A and B), encoded by two different genes, have been identified in mouse, human, rat and cattle cells [5]-[12]. The genes encoding STAT5A and STAT5B are derived from a single ancestral gene [13]; they are highly homologous, being ~90% identical in coding sequence. The two isoforms differ by few amino acids in the carboxylic end of the protein molecule [14]. Moreover, STAT5A and STAT5B show differences both in their DNA binding specificities [15], [16] and with respect to their tissue distribution [7], [8]. In cattle the STAT5A gene has been assigned to chromosome 19q17 and consists of 19 exons encoding 794 amino acids chain [13]. The STAT locus also contains STAT3 and STAT5B genes [13], [17].

Several nucleotide sequence polymorphisms of the bovine STAT5A gene (GenBank AJ242522 and AJ237937) have been detected: McCracken et al. [18] found TG repeats of different length within intron 12; Antoniou et al. [19] described two SSCP variants of the gene fragment that encodes the SH2 domain in bovine STAT5A protein; Brym et al. [20] reported a new SNP (A/G) located in intron 9 at position 9501. Khatib et al. [21] studied many SNPs in STAT5A gene and their association with embryonic survival and milk composition. Flisikowski and Zwierzchowski [22] reported a new single nucleotide polymorphism in exon 7 of the bovine STAT5A gene also investigated by other authors [23]-[27]. Moreover, Flisikowski and Zwierzchowski [28] described the substitution  $T \rightarrow C$  at position 12743 within exon 16, which changed the amino acid sequence (Val $\rightarrow$ Ala at position 686) and a deletion of CCT in intron 15, also investigated by He et al. [29] in Holstein cows. The SNP  $T \rightarrow C$  at position 12743 was shown to modify the DNA-binding properties of the protein. In the DNA-protein binding assays (electrophoretic mobility shift assay - EMSA), nuclear proteins derived from CC genotype animals always

Manuscript received January 15, 2013; revised March 22, 2013.

M. Selvaggi, V. Tufarelli, F. Pinto, A. Dambrosio, and C. Dario are with the Department DETO-Section of Veterinary Science and Animal Production, University of Bari "Aldo Moro", Italy (e-mail: maria.selvaggi@uniba.it).

G. Centoducati is with the Department of Soil, Plant and Food, University of Bari "Aldo Moro", Italy.

M. P. Santacroce is with the Department of Veterinary Medicine, University of Bari "Aldo Moro", Italy.

showed less DNA protein complexes than those of a TT animals [30].

In this study, the substitution  $T \rightarrow C$  at position 12743 within exon 16 was investigated with PCR-RFLP method in Jersey cows. There is an increasing interest for the Jersey breed that is mainly due to the characteristics of milk produced: it is excellent in terms of quality for the high fat, protein and calcium content and the low non-protein nitrogen content. Moreover, Jersey cattle ability to adapt to different breeding types and to different environmental condition contributed greatly to the breed's current success worldwide. In Italy, over the last years there has been a fairly good increase in the number of Jersey breeders even if most of the Jersey cows are bred in mixed herds with Holsteins or other breeds. That being so, the purposes of this investigation were to determine the frequencies of the variant alleles and the genotypes of the SNP in exon 16 of the bovine STAT5A gene in a sample of Jersey cows and to verify the association between this SNP and some milk production traits.

## II. MATERIALS AND METHODS

A total of ninety-five unrelated cows belonging to Jersey breed were included in the study. The animals were all primiparous and were milked twice a day. The cows, calved in two different seasons, belonged to 3 different farms located in southern Italy. They were fed with the same lactation diet, according to the energy recommendations for lactating cows, and they had free access to water.

Individual blood samples for DNA genotyping were collected from all cows on K<sub>3</sub>-EDTA tubes and stored at -25 °C. Genomic DNA was isolated from whole blood using ZR Genomic DNA II Kit<sup>TM</sup> (Zymo Research). After genomic DNA isolation, all the samples were genotyped for the gene polymorphism in exon 16 of STAT5A gene as previously described by Flisikowski et al. [30].

The following PCR primers were used STAT5A F -5'-AGC CCT ACA GCT CCA ATC CT-3' and STAT5A\_R - 5'-GGG TGT ACC CGC TGC TTA G-3 to amplify a 281-bp PCR fragment harboring parts of intron 15 and exon 16 of the STAT5A gene. The polymerase chain reactions (PCR) were performed using a PCR-mix with: each primers at a final concentration of 2 pmol/µl, 1 U Taq polymerase (SIGMA), 1  $\mu$ l Taq polymerase buffer, dNTPs of 2.0 mM/ $\mu$ l, ca. 100 ng of genomic DNA, and H2O up to 10 µl. The following PCR protocol was used: 1 min at 94 °C, 1 min at 61 °C, and 1 min at 72 °C – 34 cycles. The yield and specificity of the PCR reactions were both evaluated by electrophoresis of the amplified fragments in 2% agarose gel stained with ethidium bromide in TBE buffer. The PCR products were digested in 10-µl aliquots with 10 U of MsII restriction nuclease (BioLabs, New England, USA) for 3 hours at 37 °C. The restriction fragments were subjected to electrophoresis in 2% agarose/ethidium bromide gels in TBE buffer. The gels were examined under UV light.

The STAT5A allele frequencies were calculated by simple allele counting [31]. The polymorphism was tested for deviation from Hardy-Weinberg equilibrium (HWE) by comparing the observed and expected genotype frequencies using the  $\chi^2$  test. Data for a 305-day milk production

including milk yield (MY), protein (PC) and fat (FC) content were obtained from the local breeder association; fat and protein yielded (FY and PY, respectively) were calculated.

Effects of polymorphic variants of the STAT5A gene on milk production traits were analyzed using the GLM procedure of SAS [32] according to the following statistical model:

$$Y_{ijkl} = \mu + G_i + M_j + F_k + e_{ijkl}$$

where:  $Y_{ijkl}$  is the analysed trait of each cow;  $\mu$  is the overall mean;  $G_i$  is the fixed effect of the i<sup>th</sup> genotype (1, 2);  $M_j$  is the fixed effect of  $j^{th}$  season of calving (1, 2);  $F_k$  is the fixed effect of  $k^{th}$  farm (1,...,3);  $e_{ijkl}$  is the random error. Due to the low number of CC cows found in the population, this genotype was not included in the statistical analysis; in fact the number of CC cows is not enough to provide an accurate statistical analysis.

## III. RESULTS AND DISCUSSION

The 281-bp fragment contains two *MsI* restriction sites, only one of these appears to be polymorphic. The T $\rightarrow$ C substitution deletes one cutting site of *MsI*. All the three possible genotypes were identified. The observed frequencies of C and T alleles were 0.147 and 0.853 respectively being quite similar to those reported in other breeds (see Table I). The TT genotype was the most frequent in the studied population (73.68%) followed by TC (23.16%); only three animals were genotypes as CC (3.16%). As reported in Table II, the distribution of the genotypes was kept in Hardy-Weinberg equilibrium: the calculated  $\chi^2$  value was 0.59 (*P*=0.45; d.f.=1).

TABLE I: FREQUENCIES OF C AND T ALLELES IN JERSEY BREED AND IN DIFFERENT CATTLE BREEDS AS REPORTED BY OTHER AUTHORS. ALLELE FREQUENCIES ARE SHOWN IN DECREASING ORDER FOR THE T ALLELE

			LELIC UENCIES	
Breed	No.	T T	C	REFERENCES
Simmental	11	0.909	0.091	Flisikowski et al. [30]
Aberdeen Angus	10	0.900	0.100	Flisikowski et al. [30]
Hereford	16	0.875	0.125	Flisikowski et al. [30]
Jersey	95	0.853	0.147	Present work
Polish Friesian	150	0.850	0.150	Flisikowski et al. [33]
Limousine	16	0.812	0.188	Flisikowski et al. [30]
Charolaise	18	0.805	0.195	Flisikowski et al. [30]
Polish Friesian	37	0.756	0.244	Flisikowski et al. [30]

TABLE II: OBSERVED AND EXPECTED NUMBERS AND PERCENTAGES (IN BRACKETS) OF STAT5A GENOTYPES DETECTED BY MSLI RFLP ANALYSIS AND ALLELE FREQUENCIES IN THE SAMPLE OF JERSEY COWS

	STAT5A/MslI GENOTYPE				
NUMBER	TT	TC	CC		
OBSERVED	70 (73.68%)	22 (23.16%)	3 (3.16%)		
EXPECTED	69.06 (72.70%)	23.87 (25.13%)	2.06 (2.17%)		
$V^2 = 0.50 \text{ P} = 0.44$					

 $X^2 = 0.59 P = 0.44$ 

Data reported in Table III show the effects of the STAT5A/*MsI*I polymorphism on milk production traits. No significant differences between the TT and TC genotypes were found concerning MY (5888.20 vs. 5896.74 kg), FY (261.81 vs. 242.54 kg), PC (3.77 vs. 3.79%) and PY (219.45 vs. 223.17 kg for TT and CT respectively). On the other side, the difference concerning the fat content of milk produced by cows belonging to TC and TT groups was found significant at the statistical analysis: in particular, milk from TT animals had a higher fat content in comparison with that of TC ones (4.55 vs. 4.14%, respectively; P<0.05).

TABLE III: MEANS AND STANDARD ERROR OF MILK PRODUCTION TRAITS IN JERSEY COWS WITH DIFFERENT STAT5A/MSLI GENOTYPES

Production Traits	Genotypes		
Troduction Traits	tt	tc	
Milk yield (MY) (kg)	5888.20±96.63	5896.74±141.33	
Fat (FC) (%)	4.55±0.04a	4.14±0.10b	
Fat (FY) (kg)	261.81±2.84	242.54±6.33	
Protein (PC) (%)	3.77±0.02	3.79±0.06	
Protein (PY) (kg)	219.45±1.49	223.17±4.78	
<i>a</i> , <i>b</i> = <i>P</i> <0.05			

Even if, on the basis of these results, it is possible to

suppose a positive effect of the T allele on milk fat content, the low number of CC cows in the studied population does not permit the evaluation of the performances of this genotype and a real comparison among genotypes. However, the reason why the frequency of CC individuals in Jersey breed is very low may be indirectly due to the selective pressure.

In a previous study conducted on Polish Friesian cows, Flisikowski *et al.* [33] investigated the association among the STAT5A/*Msl*I polymorphism and some milk production traits finding results that partially agree with those reported in the present paper. In particular, they found no relationship between the studied SNP and protein and fat content of milk. On the other side, the same authors observed that cows carrying TC genotype produced daily significantly ( $P \le 0.05$ ) more milk and fat corrected milk (FCM) than TT homozygotes, with higher ( $P \le 0.05$ ) content of lactose. Moreover, the daily yield of value corrected milk (VCM), milk total solids, solids-non-fat, protein, and lactose was higher ( $P \le 0.01$ ) in cows with TC if compared to those genotyped as TT.

However, taking into account the relatively small size of the studied population, the present results should be interpreted as an association between the investigated SNP and milk traits in this population. In order to confirm these results, further investigations, including also other breeds, are necessary to better clarify the role of this SNP on production traits in cattle.

#### REFERENCES

- J. E. Darnell Jr, I. M. Kerr, and G. R. Stark, "JAK-STAT pathways and transcriptional activation in response to IFNs and other extracellular signalling proteins," *Science*, vol. 264, pp. 1415-1421, June 1994.
- [2] C. Schindler and J. E. Darnell Jr., "Transcriptional responses to polypeptide ligands. The JAK-STAT pathway," *Annual Review of Biochemistry*, vol. 64, pp. 621-651, July 1995.

- [3] H. Wakao, F. Gouilleux, and B. Groner, "Mammary gland factor (MGF) is a novel member of the cytokine regulated transcription factor gene family and confers the prolactin response," *EMBO Journal*, vol. 13, pp. 2182-219, May 1994.
- [4] L. S. Argetsinger and C. Carter-Su, "Growth hormone signalling mechanisms: involvement of the tyrosine kinase JAK2," *Hormone Research*, vol. 45, pp. 22-24, 1996.
- [5] J. Hou, U. Schindler, W. J. Henzel, S. C. Wong, and S. L. McKnight, "Identification and purification of human Stat proteins activated in response to interleukin-2," *Immunity*, vol. 2, pp. 321-329, April 1995.
- [6] A. V. Kazansky, B. Raught, S. M. Lindsey, Y. F. Wang, and J. M. Rosen, "Regulation of mammary gland factor/STAT5A during mammary gland development," *Molecular Endocrinology*, vol. 9, pp. 1598-1609, November 1995.
- [7] X. Liu, G. W. Robinson, F. Gouilleux, B. Groner, and L. Hennighausen, "Cloning and expression of Stat5 and an additional homologue (Stat5b) involved in prolactin signal transduction in mouse mammary tissue," in *Proc. the National Academy of Sciences USA*, vol. 92, pp. 8831-8835, September 1995.
- [8] A. L. Mui, H. Wakao, A. M. O'Farrel, N. Harada, and A. Miyajima, "Interleukin-3, granulocyte-macrophage colony stimulating factor and interleukin-5 transduce signals through two STAT5 homologs," *EMBO Journal*, vol. 14, pp. 1166-1175, March 1995.
- [9] J. A. Ripperger, S. Fritz, K. Richter, G. M. Hocke, F. Lottspeich, and G. H. Fey, "Transcription factors Stat3 and Stat5b are present in rat liver nuclei late in an acute phase response and bind interleukin-6 response elements," *Journal of Biological Chemistry*, vol. 270, pp. 29998-30006, December 1995.
- [10] J. X. Lin, J. Mietz, W. S. Modi, S. John, and W. J. Leonard, "Cloning of human STAT5B. Reconstitution of interleukin-2-induced STAT5A and STAT5B DNA binding activity in COS-7 cells," *Journal of Biological Chemistry*, vol. 271, pp. 10738-10744, May 1996.
- [11] C. M. Silva, H. Lu, and R. N. Day, "Characterization and cloning of STAT5 from IM-9 cells and its activation by growth hormone," *Molecular Endocrinology*, vol. 10, pp. 508-518, May 1996.
- [12] T. Goldammer, L. Meyer, H. Seyfert, R. M. Brunner, and M. Schwerin, "STAT5A encoding gene maps to chromosome 19 in cattle and goat and chromosome 11 in sheep," *Mammalian Genome*, vol. 8, pp. 705–706, September 1997.
- [13] H. Seyfert, C. Pitra, L. Meyer, R. M. Brunner, T. T. Wheeler, A. Molenaar, J. Y. McCracken, J, Herrmann, H. Thiesen, and M. Schwerin, "Molecular characterization of STAT5A- and STAT5B-encoding genes reveals extended intragenic sequence homogeneity in cattle and mouse and different degrees of divergent evolution of various domains," *Journal of Molecular Evolution*, vol. 50, pp. 550-561, June 2000.
- [14] R. Moriggl, V. Gouilleux-Gruart, R. Jahne, S. Berchtold, C. Gartmann, X. Liu, L. Hennighausen, A. Sotiropoulos, B. Groner, and F. Gouilleux, "Deletion of the carboxyl-terminal transactivation domain of MGF-Stat5 results in sustained DNA binding and a dominant negative phenotype," *Molecular and Cellular Biology*, vol. 16, pp. 5691-5700, October 1996.
- [15] C. Boucheron, S. Dumon, S. C. Santos, R. Moriggl, L. Hennighausen, S. Gisselbrecht, and F. Gouilleux, "A single amino acid in the DNA binding regions of STAT5A and STAT5B confers distinct DNA binding specificities," *Journal of Biological Chemistry*, vol. 273, pp. 33936-33941, December 1998.
- [16] F. Verdier, S. Chretien, O. Muller, P. Varlet, A. Yoshimura, S. Gisselbrecht, C. Lacombe, and P. Mayeux, "Proteasomes regulate erythropoietin receptor and signal transducer and activator of transcription 5 (STAT5) activation. Possible involvement of the ubiquitinated Cis protein," *Journal of Biological Chemistry*, vol. 273, pp. 28185-28190, October 1998.
- [17] A. Moleenar, T. T. Wheeler, J. Y. McCracken, and H. Seyfert, "The STAT3-encoding gene resides within the 40 kbp gap between the STAT5A- and STAT5B-encoding genes in cattle," *Animal Genetics*, vol. 31, pp. 333–346, June 2000.
- [18] J. Y. McCracken, A. J. Molenaar, R. J. Snell, H. W. Davey, and R. J. Wilkins, "A polymorphic TG repeat present within the bovine STAT5A gene," *Animal Genetics*, vol. 28, pp. 453-464, August 1997.
- [19] E. Antoniou, B. J. Hirts, M. Grosz, and J. Skidmorec, "A single strand conformation polymorphism in the bovine gene STAT5A," *Animal Genetics*, vol. 30, pp. 232, December 1999.
- [20] P. Brym, S. Kamiński, and A. Ruść, "A New SSCP polymorphism within bovine STAT5A gene and its associations with milk performance traits in Black-and-White and Jersey cattle," *Journal of Applied Genetics*, vol. 45, pp. 445-452, April 2004.

- [21] H. Khatib, R. L. Monson, V. Schutzkus, D. M. Kohl, G: J. M. Rosa, and J. J. Rutledge, "Mutations in the *STAT5A* gene are associated with embryonic survival and milk composition in cattle," *Journal of Dairy Science*, vol. 91, pp. 784-793, February 2008.
- [22] K. Flisikowski, and L. Zwierzchowski, "Single-strand conformation polymorphism within exon 7 of the bovine STAT5A gene," *Animal Science Papers and Reports*, vol. 20, pp. 133-137, February 2002.
- [23] C. Dario, M. Dario, F. Ciotola, V. Peretti, D. Carnicella, and M. Selvaggi, "Analysis of STAT5A/AvaI gene polymorphism in four Italian cattle breeds," *Biochemical Genetics*, vol. 47, pp. 671-679, June 2009.
- [24] C. Dario, M. Selvaggi, D. Carnicella, and G. Bufano, "STAT5A/AvaI polymorphism in Podolica bulls and its effect on growth performances traits," *Livestock Science*, vol. 123, pp. 83-87, July 2009.
- [25] M. Selvaggi, C. Dario, G. Normanno, G. V. Celano, and M. Dario, "Genetic polymorphism of STAT5A protein: relationships with production traits and milk composition in Italian Brown cattle," *Journal of Dairy Research*, vol. 76, pp. 441-445, July 2009.
- [26] M. Sadeghi, M. M. Shahrbabak, G. R. Mianji, and A. N. Javaremi, "Polymorphism at locus of STAT5A and its association with breeding values of milk production traits in Iranian Holstein bulls," *Livestock Science*, vol. 123, pp. 97-100, July 2009.
- [27] M. Selvaggi and C. Dario, "Study on the STAT5A/AvaI polymorphism in Jersey cows and association with milk production traits," *Molecular Biology Reports*, vol. 38, pp. 5387-5392, March 2011.
- [28] K. Flisikowski and L. Zwierzchowski, "Polymerase chain reaction-heteroduplex (PCR-HD) polymorphism within the bovine

STAT5A gene," *Journal of Applied Genetics*, vol. 44, pp. 185–189, February 2003.

- [29] X. He, M. X. Chu, L. Qiao, J. N. He, P. Q. Wang, T. Feng, R. Di, G. L. Cao, L. Fang, and Y. F. An, "Polymorphisms of STAT5A gene and their association with milk production traits in Holstein cows," *Molecular Biology Reports*, vol. 39, pp. 2901-2907, June 2012.
- [30] K. Flisikowski, M. Szymanowska, and L. Zwierzchowski, "The DNA binding capacity of genetic variants of the bovine STAT5A transcription factor," *Cellular and Molecular Biology Letters*, vol. 8, pp. 831–840, March 2003.
- [31] D. S. Falconer and T. F. C. Mackay, *Introduction to Quantitative Genetics*, 4th edition Longman Group Ltd, Essex, 1996.
- [32] SAS– SAS/STATTM, Guide for Personal Computers, version 8.1, Ed. NC: SAS Istitute Inc., Cary, USA, 1999/2000.
- [33] K. Flisikowski, N. Strzałkowska, K. Słoniewski, J. Krzyżewki, and L. Zwierzchowski, "Association of a sequence nucleotide polymorphism in exon 16 of the STAT5A gene with milk production traits in Polish Black-and-White (Polish Friesian) cows," *Animal Science Papers and Reports*, vol. 22, pp. 515-522, April 2004.

**Maria Selvaggi** was born in Italy and received her master's degree in Veterinary Medicine at University of Bari "Aldo Moro". She has earned her Ph.D in "Hygiene, Public Health and Food Control" at University of Bari. At present, she is working as researcher at Department DETO- Section of Veterinary Science and Animal Production- in the field of Animal Genetics.